

Enzymatic Synthesis of Bicyclic γ -Lactams using Clavaminic Acid Synthase

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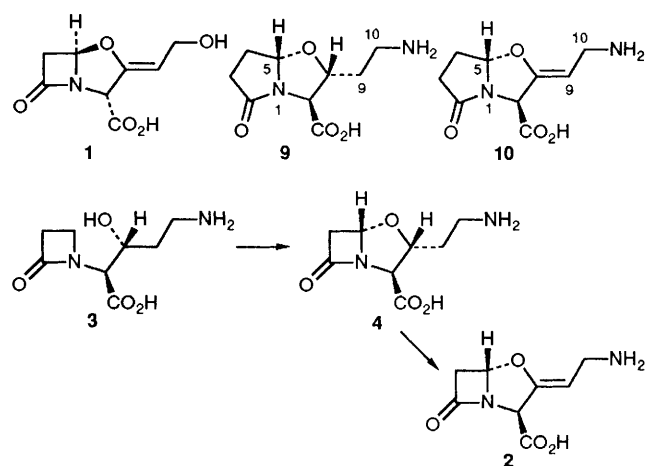
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Incubation of a γ -lactam analogue of proclavaminic acid with clavaminic acid synthase, gave two novel bicyclic γ -lactams.

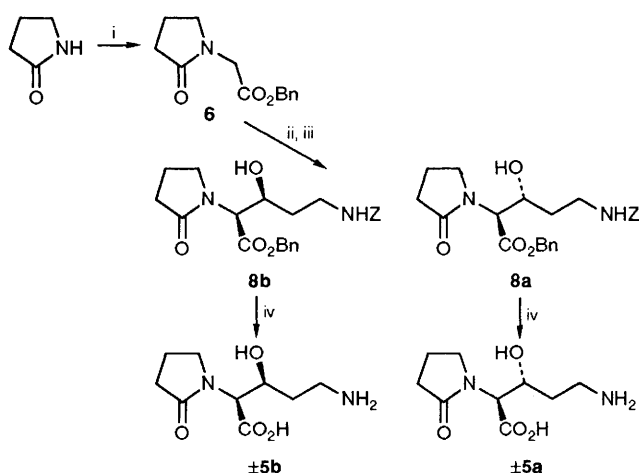
Clavulanic acid **1** is a commercially important β -lactamase inhibitor.¹ Studies on the biosynthesis of **1** by the SmithKline Beecham group resulted in the isolation of clavaminic acid **2** which was shown to be a precursor of clavulanic acid **1**. Furthermore a monocyclic β -lactam, proclavaminic acid **3** was also isolated, which in turn was shown to be a precursor of clavaminic acid **2**.^{2,3} The enzyme responsible for the catalytic oxidative cyclisation and desaturation of proclavaminic acid **1** to give clavaminic acid **2** has been purified from *Streptomyces clavuligerus* and has been partially characterised.⁴ The enzyme, clavaminic acid synthase (CAS) was shown to require dioxygen and α -ketoglutarate as cosubstrates and to be dependent on iron(II) for activity. Hence, it belongs to the

family of 2-oxo acid dependent dioxygenases, some of which also catalyse key steps in the biosynthesis of other β -lactam microbial metabolites. The gene encoding CAS has been identified, cloned and CAS has been over expressed in *Escherichia coli*.⁵ Recent *in vitro* experiments using CAS have determined that cyclisation precedes desaturation and that the saturated clavam **4** is an intermediate in the conversion of **3** into **2** (Scheme 1).⁶

Extensive substrate analogue studies on oxygenases involved in the biosynthesis of the penicillins and cephalosporins have established that they have a lax specificity with regard to unnatural substrates and have resulted in mechanistic proposals.⁷ We speculated that a similar approach



Scheme 1

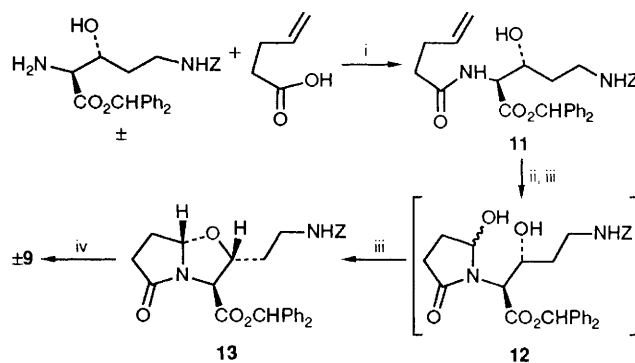


Scheme 2 Z = PhCH₂OCO; Bn = PhCH₂ Reagents i, NaH, dimethylformamide (DMF), BrCH₂CO₂Bn (77%); ii, (Me₃Si)₂NLi, tetrahydrofuran (THF), -78 °C; iii, OHC(CH₂)₂NH₂ **7** then H₃O⁺ (30% for **8a**); iv, H₂, 10% Pd/C, EtOH-H₂O (1:1) (65%, in each case)

may prove fruitful both in the study of CAS and its *in vitro* use as a synthetic reagent. Herein, we report the enzymatic synthesis of bicyclic γ -lactams exploiting CAS, from a monocyclic γ -lactam substrate \pm **5a**.

The requisite substrate was synthesised in racemic form using an extension of methodology developed for the synthesis of proclavaminc acid **3**.³ Thus, the pyrrolidinone **6** was selectively deprotonated on the exocyclic methylene group and the resultant enolate quenched with the aldehyde **7** to give a mixture of *threo* **8a** and *erythro* **8b** alcohols **8a**:**8b**, 3:1, in moderate yield, which were separated by HPLC. Deprotection gave the desired *threo* **5a** and *erythro* **5b** compounds (Scheme 2).

Analysis by ¹H NMR (500 MHz) of incubations of **5a** with CAS[†] and the appropriate cofactors⁴ indicated the presence of



Scheme 3 Reagents: i, 2-ethoxy-1-ethoxycarbonyl-1,2-dihydroquinoline, CH₂Cl₂, (81%); ii, NaIO₄, OsO₄ (cat.), THF, H₂O (3:1); iii, CH₂Cl₂, CF₃CO₂H (cat.) (30% from **11** plus ca. 25% recovered starting material); iv, H₂, 10% Pd/C, THF, H₂O (1:1) (85%)

two new bicyclic materials. These were purified by reversed-phase HPLC and assigned as the saturated bicyclic γ -lactam **9** and the γ -lactam analogue of clavamic acid **10**.[‡] For **9**: HPLC Waters Bondapak amine column, 0.015% HCO₂H in 95% H₂O-5% MeOH; δ_{H} (500 MHz, D₂O) 1.74-1.93 (2 H, m, 9-H and 6-H), 2.02-2.09 (1 H, m, 9-H), 2.24-2.32 (1 H, m, 6-H), 2.34-2.40 (1 H, m, 7-H), 2.57-2.75 (1 H, m, 7-H), 3.05 (2 H, *ca.* t, *J* 7 Hz, 2 \times 10-H), 3.92 (1 H, d, *J*, 6.5 Hz, 2-H), 4.10-4.15 (1 H, m, 3-H) and 5.25 (1 H, dd, *J* 6.5 and 3 Hz, 5-H); A 2D COSY (correlation spectroscopy) experiment was consistent with the connectivity as indicated; *m/z* (electrospray) 215 (MH⁺). For **10**: (i) HPLC as for **9**, then (ii) HPLC octadecylsilane reversed-phase column, 25 mmol dm⁻³ NH₄HCO₃ in H₂O; δ_{H} (500 MHz, D₂O) 2.06-2.11 (1 H, m, 6-H), 2.42-2.73 (2 H, m, 6-H and 7-H), 2.7 (1 H, dd, *J* 16 and 8 Hz, 7-H), 3.46-3.58 (2 H, m, 2 \times 10-H), 4.75 (1 H, dt, *J* 7 and 1 Hz, 9-H), 4.86 (1 H, *ca.* s, 2-H) and 5.66 (1 H, dd, *J* 6.5 and 3 Hz, 5-H); A 2D COSY experiment was consistent with the connectivity as indicated; *m/z* (electrospray) 213 (MH⁺). The relative stereochemical assignment of **9** was initially made on the basis of NOE (nuclear Overhauser effect) experiments [selected data only: irradiation at 5.25 (5-H) enhanced signals at 4.10-4.15 (3-H, 8%) and 2.24-2.32 (6-H, 9%); irradiation at 4.10-4.15 (3-H) enhanced signals at 5.25 (5-H, 8%), 3.05 (2 \times 10-H, 4%), 2.02-2.09 (9-H, 4%) and 1.74-1.93 (9-H and 6-H 3%); irradiation at 3.92 (2-H) enhanced signals at 3.05 (2 \times 10-H, 4%), 2.02-2.09 (9-H, 4%) and 1.74-1.93 (6-H and 9-H, 6%)]. The *erythro* material \pm **5b** was found not to give any bicyclic lactams upon incubation with CAS. Neither **9** nor **10** showed significant biological activity either as antibacterial agents or as β -lactamase inhibitors.

The structural assignments and relative stereochemistry of the new products were further substantiated by synthesis of \pm **9**. The strategy employed used methodology previously developed for the synthesis of γ -lactam analogues of the oxa-penam.⁸ Thus, the amide **11**, synthesised from diprotected racemic *threo*- β -hydroxyornithine⁹ and pent-4-enoic acid, was oxidised and cyclised to give a single isolated bicyclic lactam **13**, *via* the epimeric alcohols **12**. Lactam **13** was deprotected to give the desired γ -lactam \pm **9** (Scheme 3), which was shown by doping experiments to be indistinguishable by ¹H NMR (500 MHz) from the biosynthetic sample. Furthermore, incubation of the racemic synthetic **9** with CAS also gave **10**, confirming **9** to be an intermediate between **5a** and **10**.

This synthesis of bicyclic γ -lactams utilising CAS is the first report of the transformation of a novel substrate by CAS and indicates that, like isopenicillin N synthase, CAS may have a relatively relaxed specificity towards unnatural substrates.

[†] The CAS used in this study was obtained from both recombinant and wild-type sources. Full details will be published elsewhere. In a typical incubation protocol, 2 mg of \pm **5a** were incubated with 0.2 IU partially purified 'recombinant' CAS (specific activity: 2.5 IU mg⁻¹) according to previously published protocols,⁶ to give after HPLC purification **9** (*ca.* 15%) and **10** (*ca.* 15%). Longer incubation times increased the ratio of **10** to **9**.

[‡] The absolute stereochemistry of products **9** and **10** was preliminarily assigned by analogy to the conversion of the natural substrate **3** to **2**.

Future studies will attempt to further exploit this propensity in investigations concerning the mechanism of CAS.

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